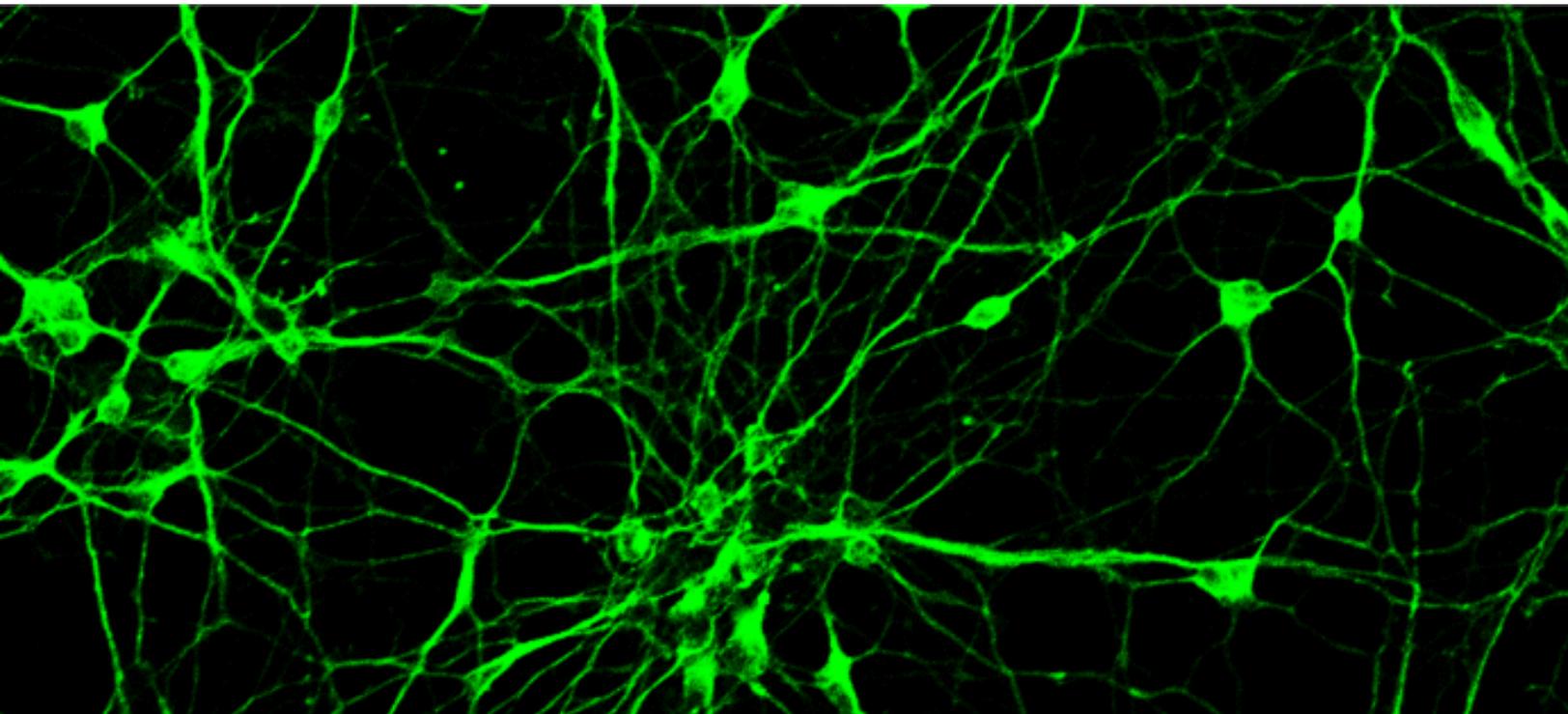




# Neurosight<sup>®</sup>-S User Guide



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## Product Information

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### Unpacking & Handling

- ▼ Upon receiving the shipment of Neurosight<sup>®</sup>-S, check whether all temperature-sensitive components are correctly frozen. If this is not the case, please contact our support team.
- ▼ Immediately transfer each of the components to the appropriate storage conditions.
- ▼ Please check the catalog number, lot number and expiry date. The basal media expiration date is the shortest (1 year) so experiments should be planned accordingly.
- ▼ The Neurosight<sup>®</sup>-S should be handled by technically qualified individuals complying with good laboratory practices, applicable laboratory regulations and the MSDS. Following the User Guide herein is recommended for best results.
- ▼ The Neurosight<sup>®</sup>-S is intended for research use only, not intended for any type of use in animals or humans.

## Components & Description

COMPONENTS	CAT#	STORAGE ON ARRIVAL
<b>Neurosight®-S Neurons</b>		Liquid Nitrogen
Cryopreserved, frozen vial		
>2 million cells	N-001	
>4 million cells	N-002	
<b>Neurosight®-S Media</b>		-20°C
Basal medium		
50 ml	NM-001	
100 ml	NM-002	
<b>Neurosight®-S Media Supplement (100X)</b>		-20°C
0.5 ml	NS-001	
1 ml	NS-002	
<b>Neurosight®-S User Guide<sup>1</sup></b>		
<b>Certificate of Analysis (CoA)</b>		
<b>MSDS<sup>2</sup></b>		

<sup>1</sup> Also available online at [www.nexel.co.kr](http://www.nexel.co.kr)

<sup>2</sup> Enclosed with shipping documents.

*Should any of the above components be missing from your shipment, please contact us at NEXEL Co. Ltd. or the distributor in your country upon which our support team will provide the necessary assistance.*

### Neurosight®-S Neurons

<b>Cell Type</b>	Human Induced Pluripotent Stem Cell (hiPSC) derived neurons
<b>Cell Line of Origin</b>	hiPSC cell line reprogrammed from commercially available normal donor fibroblast cell line
<b>Quality Control</b>	Please refer to the CoA for lot-specific information. Virus clearance & STR analysis data is available upon request.

### Neurosight®-S Media & Media Supplement

- ❖ The Neurosight®-S Media & Media Supplement need to be combined to make the Neurosight®-S Maintenance Media, after which it should be used within 1 month. DO NOT FREEZE the Neurosight®-S Maintenance Media, aliquot into smaller quantities for best results.
- ❖ The Neurosight®-S Maintenance Media is animal component and serum-free. For additional information on the composition, please contact our technical support team.
- ❖ The Neurosight®-S Maintenance Media is antibiotic and antifungal free as they are not necessary if proper conditions are kept. NEXEL does not recommend the use of such agents for accurate results but they should be used if aseptic cell culture conditions are not possible.

## Safety Precaution & User Notice



Biosafety Level: 1

For research use only, not intended for any type of use in animal or humans. Appropriate safety procedures should always be used with this material. Please refer to the MSDS for detailed instructions.

User Notice & Restrictions:

- ▼ User may use the Product (Neurosight<sup>®</sup>-S) for internal research including but not limited to screening potential drug compounds for efficacy and safety, and for the provision of such services to third parties. No other right is granted to User whether expressly, by implication, by estoppel or otherwise. In particular, the purchase of the Product does not include nor carry any right or license to use, develop or otherwise exploit the Product commercially, and no rights are conveyed to User to use the Product for any other purpose.
- ▼ User agrees to use the Product in compliance with all applicable statutes and regulations, but not to use the Product for any administration or application to humans. Moreover, User agrees not to use the Product in human subjects for human clinical use for therapeutic, diagnostic or prophylactic purposes, or in animals for veterinary use for therapeutic, diagnostic or prophylactic purposes, including but not limited to clinical applications, cell therapy, transplantation, and/or regenerative medicine without an appropriate license.
- ▼ In the case that User transfers Product to a third party, User shall convey the User Restrictions set forth herein to such third party.

# 1. Introduction

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NEXEL Co., Ltd. strives to provide high quality human neurons derived from induced pluripotent (iPS) cells using optimized proprietary protocols. The Neurosight<sup>®</sup>-S is a highly pure and electrophysiologically active population of cells, ensuring researchers get a reliable product. Thus, NEXEL hopes to help the advance of science in tissue-specific research, toxicity screening and drug discovery.

This User Guide will help you seed the Neurosight<sup>®</sup>-S at the appropriate densities to create mature neuronal networks comprising functional neurons appropriate for a variety of applications related to the electrophysiological behavior such as calcium fluxes or MEA assays and neurite outgrowth or degeneration. However, please keep in mind that the best individual results will be obtained by close observation, care and optimization from the user. Example morphology of the Neurosight<sup>®</sup>-S can be found below as a reference.

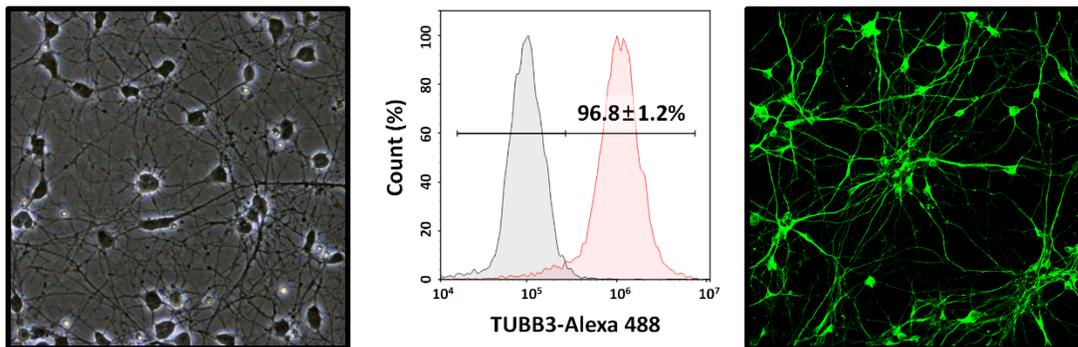


Figure 1. Neurosight<sup>®</sup>-S morphology by phase contrast microscopy and immunoassays demonstrates high purity neurons. *i) Phase contrast image at 14 DIV, ii) Flow cytometry with TUJ1 (Beta tubulin III) and iii) TUJ1 immunostaining image.*

## 2. Preparing for Cell Culture

### Required Equipment and Consumables (Not Provided)

ITEM	CAT#	VENDOR
<b>Poly-L-ornithine</b> Solution, 50 ml <sup>1,2</sup>	P4957- 50ML	Sigma
<b>Laminin</b> Solution, Variable Concentration & Volume <sup>1</sup>	354232	Corning
<b>Typical Cell Culture Equipment</b>		
Liquid Nitrogen Storage Tank		
37°C Water Bath		
Tabletop Centrifuge		
Biological Safety Cabinet with UV Lamp		
Hemocytometer or Automated Cell Counter		
Phase Contrast Microscope		
Pipettes		
Cell Culture Incubator		
<b>Typical Cell Culture Consumables</b>		
Centrifuge Tubes		
Cell Culture Plates		
Pipette Tips		
Trypan Blue		
Phosphate Buffered Saline (PBS)		

<sup>1</sup> It is possible to use materials purchased from other vendors, these are our suggestions for products which are both available world-wide and verified at NEXEL.

<sup>2</sup> Other replacements such as Poly-L-lysine or PEI which are used for neuronal cultures can replace Poly-L-ornithine.

## Preparing Neurosight®-S Media

- I. Thaw the Neurosight®-S Media by placing it at 4°C 24 hours prior to use.
- II. In a biosafety cabinet, thaw (at RT) the Neurosight®-S Media Supplement (100X) and add it to the thawed media to make Neurosight®-S Maintenance Media. Store at 4°C and away from light for up to 1 month. DO NOT FREEZE Neurosight®-S Maintenance Media.

*To avoid oxidation of the media due to air contact and repeated warming/opening, it is recommended to aliquot the media into quantities enough for 2~3 media changes.*

<b>MEDIA TYPE</b>	<b>COMPONENTS</b>
<b>Neurosight®-S Maintenance Media</b>	<b>Neurosight®-S Media Neurosight®-S Media Supplement</b>

## Preparing Cell Culture Surfaces

- I. Calculate the amount of coating media required using the following table as a reference.

CELL CULTURE PLATE	6-well (9.6 cm <sup>2</sup> )	12-well (3.8 cm <sup>2</sup> )	24-well (1.9 cm <sup>2</sup> )	48-well (1.0 cm <sup>2</sup> )	96-well (0.33 cm <sup>2</sup> )
COATING VOLUME	1 ml	500 µl	300 µl	100 µl	50 µl

- II. Coat the cell culture plates using Poly-L-ornithine solution for > 1 hour at 37°C.
- III. Prepare the Laminin coating solution to working concentrations immediately before use (at the end of 1 hour) as described in the following table. The coating solution can be kept at 4°C for a short period of time but this is not recommended.

COATING TYPE	STOCK CONCENTRATION	WORKING CONCENTRATION
Laminin	Variable, Check on Bottle	5 µg/ml

- IV. Wash the wells twice with PBS (2x the coating volume).

*It is critical to not let it dry before the first wash. After the second wash, while performing step IV, either aspirate immediately or once the Laminin solution is ready.*

- V. Pipette the correct amount of Laminin coating solution to each well you intend to use.
- VI. Gently swirl the plate and check whether all the wells are completely covered.
- VII. Incubate at 37°C for at least an hour.

## 3. Neurosight®-S Thawing and Plating

### Thawing

The Neurosight®-S can be thawed using typical cell culture thawing protocols. Here, we present NEXEL's optimized protocol and recommend our users to follow the instructions to maximize results. We strongly recommend thawing 1 vial at a time to minimize cell exposure to liquid DMSO.

- I. Calculate the amount of Neurosight®-S Maintenance Media required. For each vial, 10 ml of Maintenance Media is required for thawing. The amount required can be calculated by the number of wells; recommendations for different cell culture plates can be found below.

CELL CULTURE PLATE	6-well (9.6 cm <sup>2</sup> )	12-well (3.8 cm <sup>2</sup> )	24-well (1.9 cm <sup>2</sup> )	48-well (1.0 cm <sup>2</sup> )	96-well (0.33 cm <sup>2</sup> )
PLATING VOLUME	2 ml	1 ml	500 µl	300 µl	200 µl

- II. Warm the Neurosight®-S Maintenance Media at Room Temperature (RT, 25°C) for at least 30 mins. For each vial to thaw, aliquot 8 ml of Maintenance Media in a 15 ml centrifuge tube.
- III. Retrieve the Neurosight®-S vial(s) from the liquid nitrogen storage tank.
- IV. Submerge the vial(s) 2/3 in a 37°C water bath so that the mouth of the vial does not come in contact with the water. Constantly check how much has thawed and once ~20% remain (~3 mins), spray the vial(s) with 70% Et-OH, wipe and place it in your biosafety cabinet. Ideally, the vial(s) should have completely thawed exactly when you start step V.
- V. Open the vial(s) and transfer the contents (~1 ml) using a 1 ml pipette to the aliquoted 8 ml of Maintenance Media dropwise while gently swirling the tube.

*Dropwise pipetting while gently swirling the tube minimizes osmotic shock and maximizes mixing, which ensures high viability. Drops will remain on the surface for ~1 second and then drop towards the bottom of the tube (visible due to the DMSO content). For dropwise pipetting, simply pipette slowly into the air ~1 cm above the media surface. It should take approximately 1 min per 1 ml.*

- VI. Use 1 ml of Maintenance media to gently rinse the emptied vial and transfer dropwise to the centrifuge tube containing the cells from step V while gently swirling the tube.

- VII. Centrifuge the suspended cells at 180 x g for 3 minutes at room temperature.
- VIII. Carefully discard the supernatant.
- IX. Resuspend the cells gently using 1 ml of Plating Medium and check the cell concentration using a hemacytometer or cell counter and Trypan Blue. Immediately move on to the Plating section.

*Avoid rigorous pipetting of the cells to maximize viability. Single cell resuspension of the Neurosight<sup>®</sup>-S during thawing should easily be achieved by gently pipetting 3~4 times.*

## Plating

NEXEL recommends seeding the Neurosight<sup>®</sup>-S at a density of ~100,000 cells/cm<sup>2</sup> for most standard applications. Application specific protocols are available upon request (will be added to the website in early 2020). The best results are obtained by the User's own optimization, for which NEXEL will try to provide as much assistance as possible.

- I. Calculate the volume of Plating Media and cells required to match the correct density for the culture platform of choice. Below is a table with cell numbers.

CELL CULTURE PLATE	6-well (9.6 cm <sup>2</sup> )	12-well (3.8 cm <sup>2</sup> )	24-well (1.9 cm <sup>2</sup> )	48-well (1.0 cm <sup>2</sup> )	96-well (0.33 cm <sup>2</sup> )
PLATING VOLUME	2 ml	1 ml	500 µl	300 µl	200 µl
CELL NUMBER	960,000	380,000	190,000	100,000	33,000

*Well area (cm<sup>2</sup>) can vary between different vendors, please check with your providers for exact calculations.*

- II. Combine the volumes as calculated above. Add laminin to a final concentration of 1 µg/ml.
- III. Remove the coating solution in the cell culture plates. Avoid drying out the coated wells as much as possible.
- IV. Gently mix by pipetting and evenly distribute the appropriate volumes of cells with Plating Media.
- V. Move the cell culture plate to the incubator, shake the plate in perpendicular directions to evenly distribute the cells for attachment.
- VI. 2 days later, perform the first media change with Maintenance Media (Neurosight<sup>®</sup>-S Maintenance Step I)

## 4. Neurosight®-S Maintenance

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Starting two days after plating the cells, the media needs to be changed every two to three days. Ideally, the media should be changed at 48-hour intervals. When performing electrophysiological assays on the cells, we recommend changing the media 1 hour prior to the experiment to ensure there are enough nutrients for the cells.

- I. Warm the correct volume of Neurosight®-S Maintenance Media volume (half of the volume for plating) at Room Temperature for at least 30 mins.
- II. Immediately before media change, add Laminin to media to a final concentration of 1 µg/ml
- III. Perform a half-media change with the newly warmed media in a biosafety cabinet. Pipette softly onto the cell culture plate walls to avoid any damage to the cell culture.

*Performing half-media changes is crucial for neuronal cultures. Firstly, even the shortest contact with air can damage pure neuronal cultures and cell death will be visible within a few days. Secondly, it prevents removal of cytokines secreted by the cells themselves resulting in higher quality cell cultures. An example of half-media change is: for 1 ml of media in the well, remove ~ 400 µl and add 500 µl.*

*Neuronal cultures are very sensitive to pipetting. Both upon removal and addition of media, users should take extra care to not result in detachment of the culture.*

- IV. Place the plate back in the incubator.
- V. Repeat I to IV every 2~3 days.

We recommend performing any planned assays with the Neurosight®-S from Day 10 onwards. Long term culture over 21 days can lead to aggregation & cell detachment at high densities, we recommend careful & constant observation of the cell culture. Aggregation on its own is not always detrimental to experiment results, particularly electrophysiological assays. We do not recommend subculture/passaging of the neuronal cell cultures as mature neurons are very fragile and neurite destruction may lead to cell death.